

PGR management in the genomics era

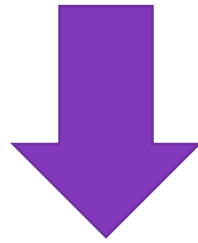
WP3

Lorenzo Barchi, Ezio Portis, Giuseppe Aprea, Jan Bartos, Jaroslav Dolezel, Giovanni Giuliano



Omic technologies represent a leap forward for the conservation, management and characterization of PGR

Current PGR management does not involve the routine use of genomic tools to trace accessions during seed regeneration or vegetative propagation



Taxonomical classification on morphological traits alone can be error-prone



Genomics-based identification should become the golden standard for the management of PGR.



Genebank accessions should have their DNAs (accessions) barcoded or sequenced

PGR *in situ* managing can benefit from the use of genomics



Many smaller genebanks, *in situ* collection holders and final users of the PGR as well as scientists working on PGR simply don't have the facilities and skills to use these technologies.

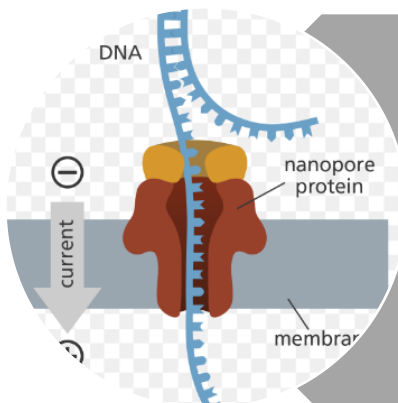
Refine and suggest protocols and methods for DNA barcoding (D3.2)

DNA (accessions) barcoding and genetic classification based on:



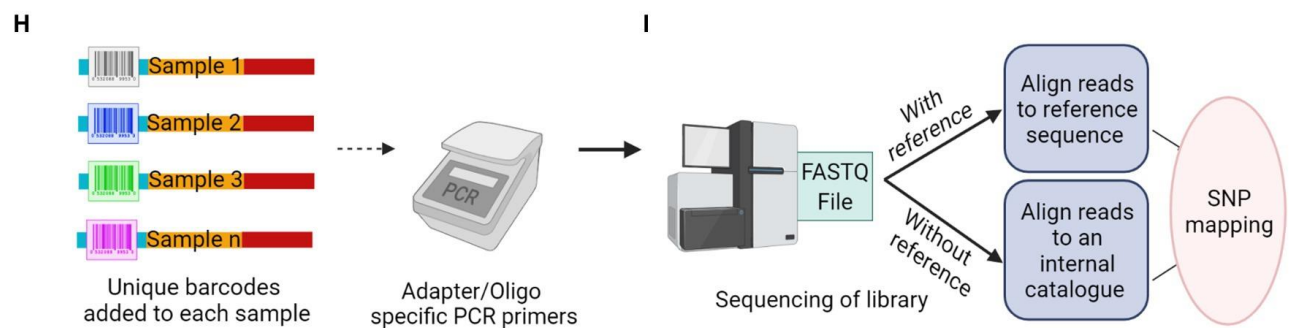
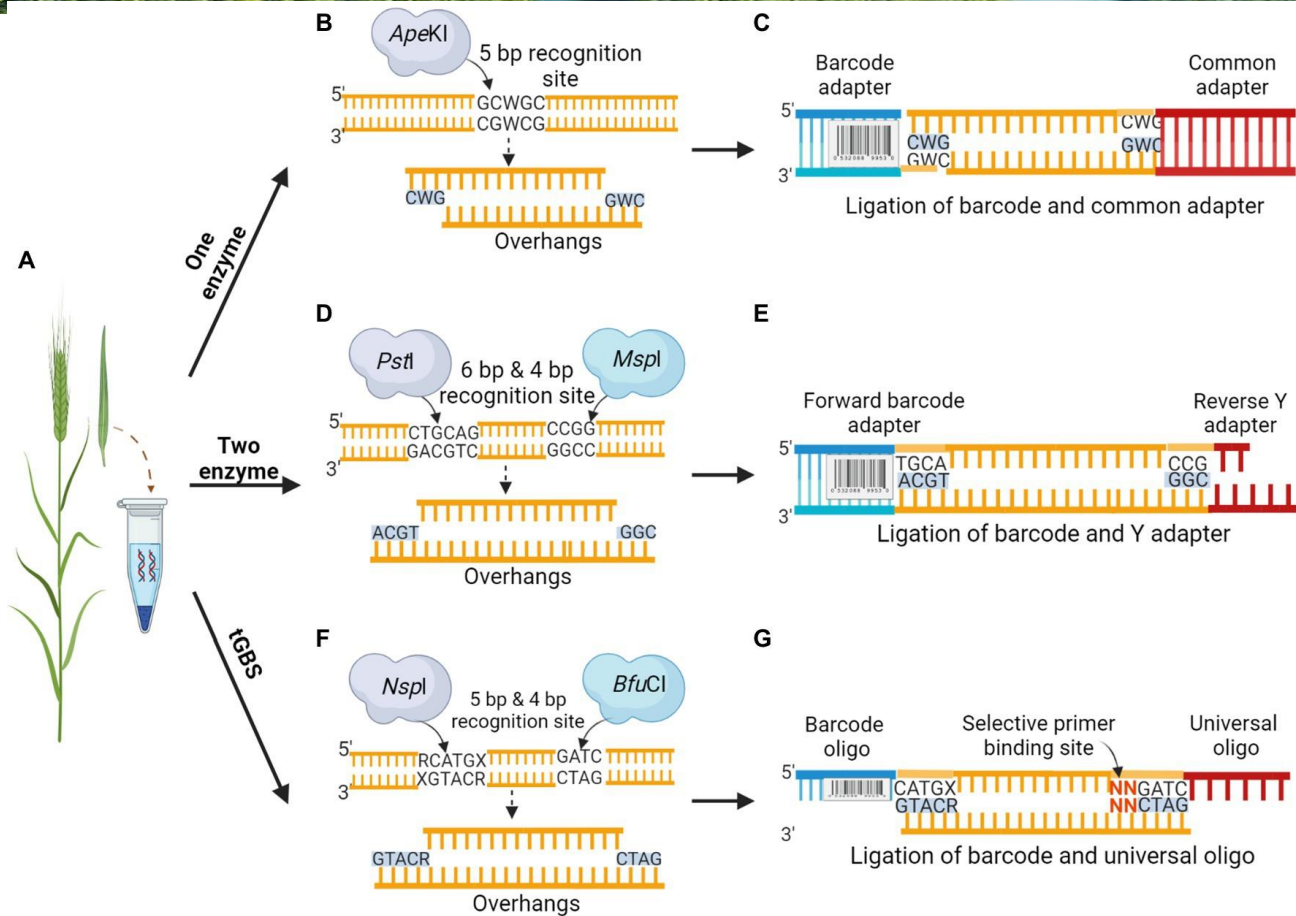
Reduced representation sequencing (RRS).

Using a set of common SNPs, it is now possible to identify thousands of polymorphism at low cost for accession identification



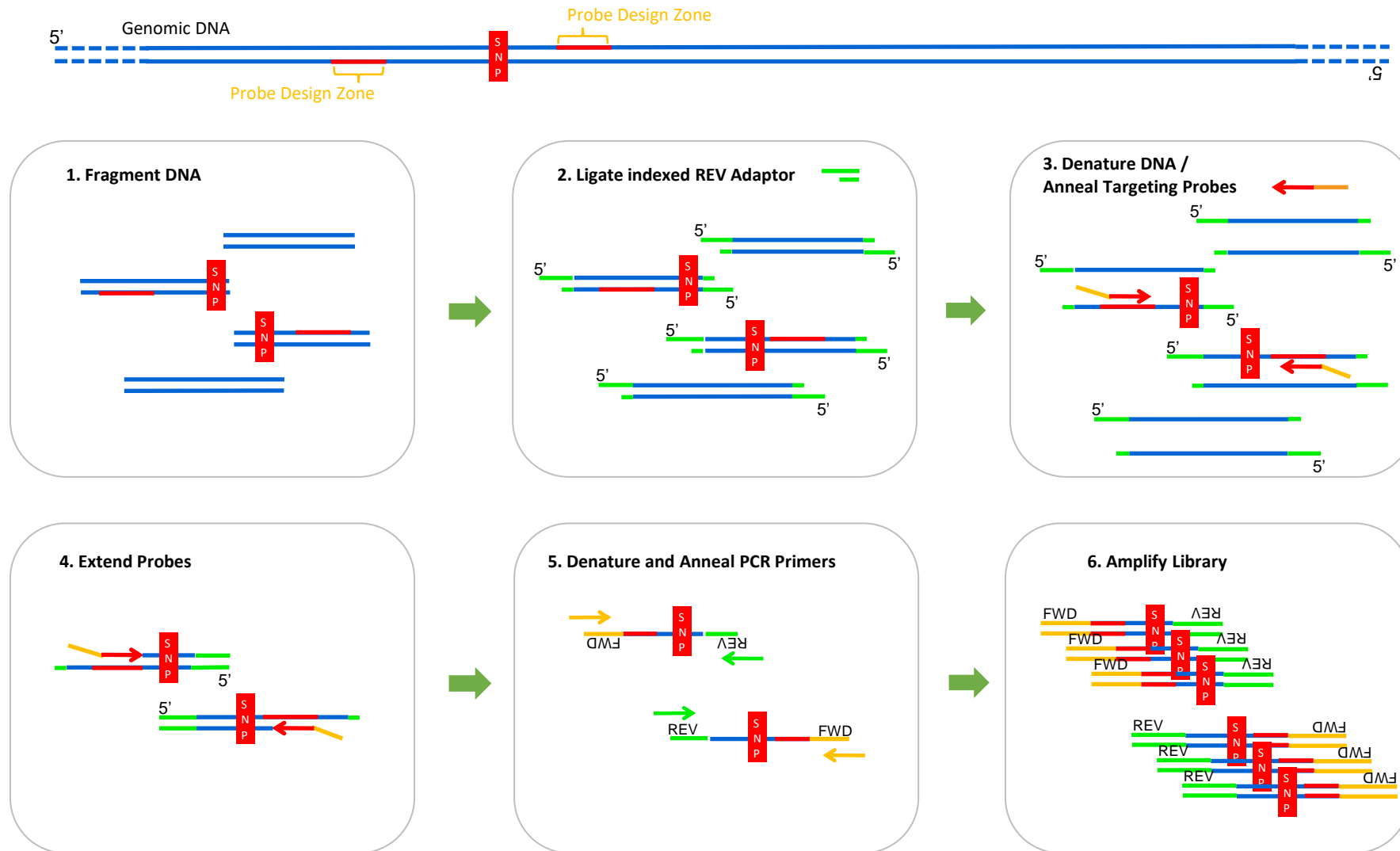
Whole genome resequencing (short reads) and chromosome scale sequencing (based on long reads)

GBS: genotyping by sequencing



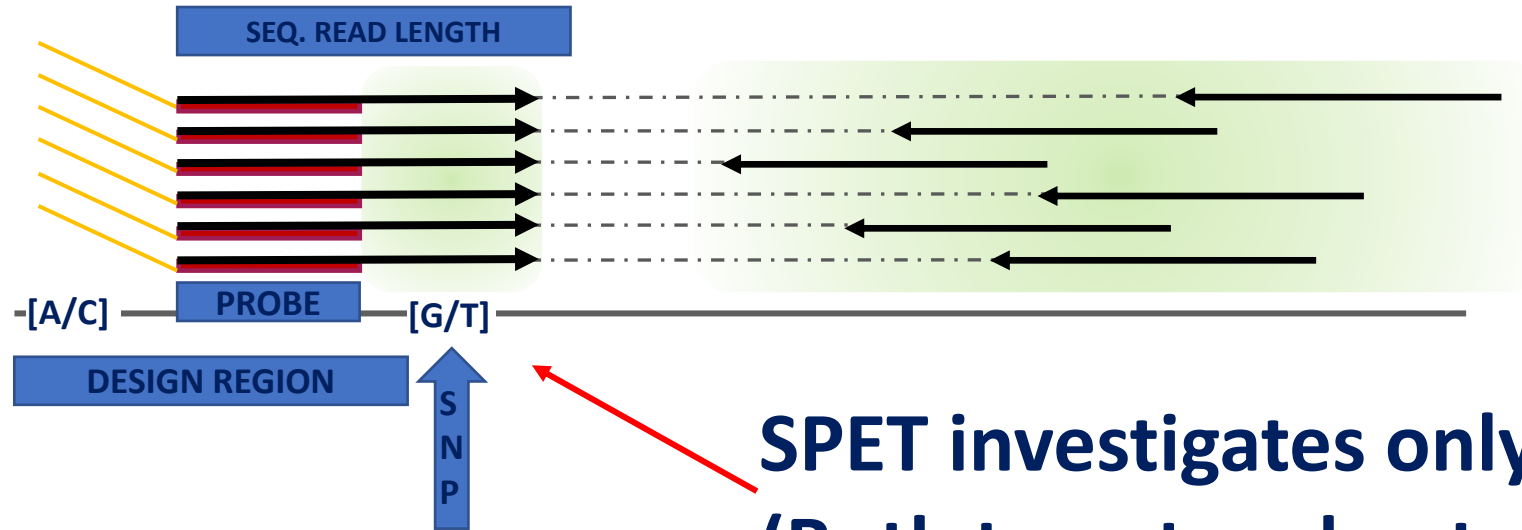
Reduced representation sequencing

Single Primer Enrichment Technology (SPET)



Reduced representation sequencing

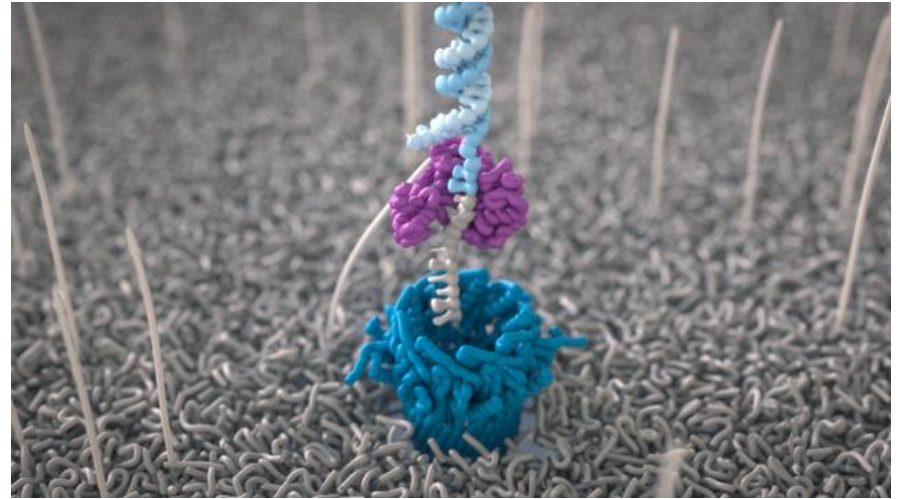
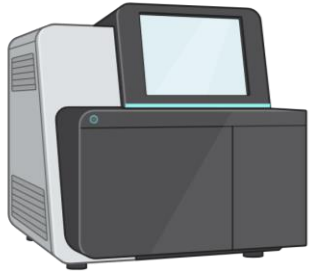
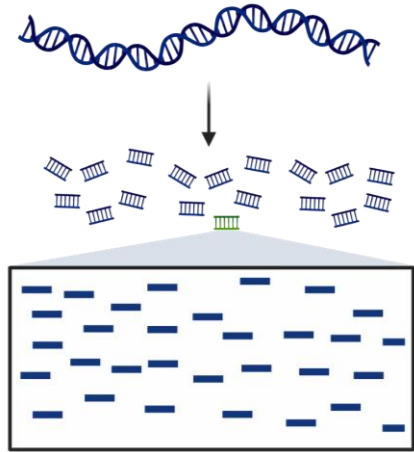
Target Enrichment Application: Genotyping By Sequencing



**SPET investigates only TARGET loci
(Both target and untargeted SNPs)**

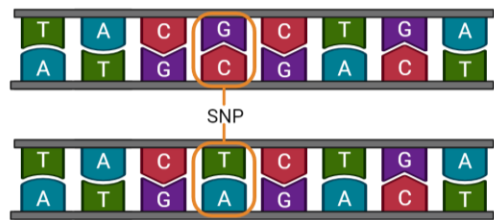
- Highly-multiplex genotyping (up to 3,092)
- Enables detection of a large number of known SNPs
- Every sequencing read is informative

WGS

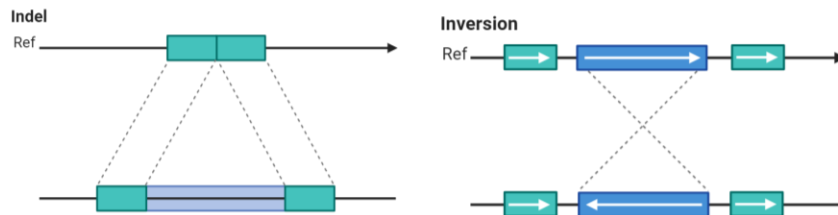


Sequencing data as well as **Pangenome** construction

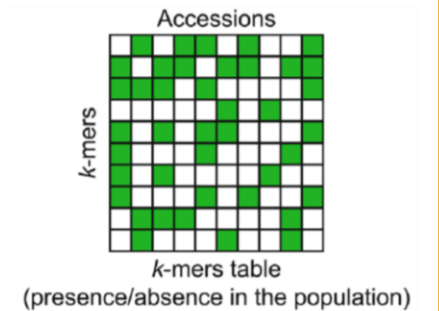
SNPs



Structural variations (SVs)



K-mers



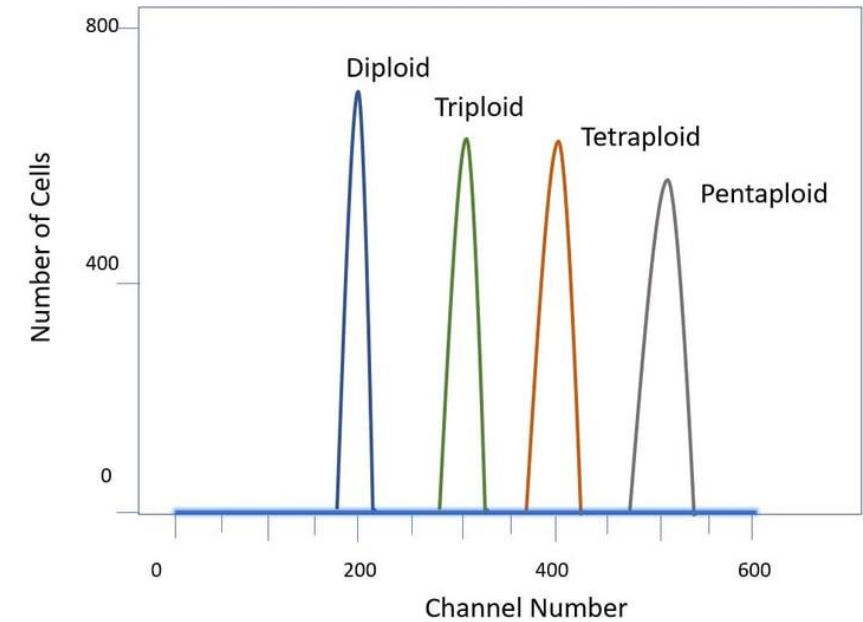
D3.2: Cytogenomics

Cytogenomics protocols for:

➔ Ploidy/aneuploidy/genome size determination (especially for clonally propagated species)

➔ Providing precious information on chromosome number, karyotype and genomic constitution of CWR (Crop Wild Relatives)

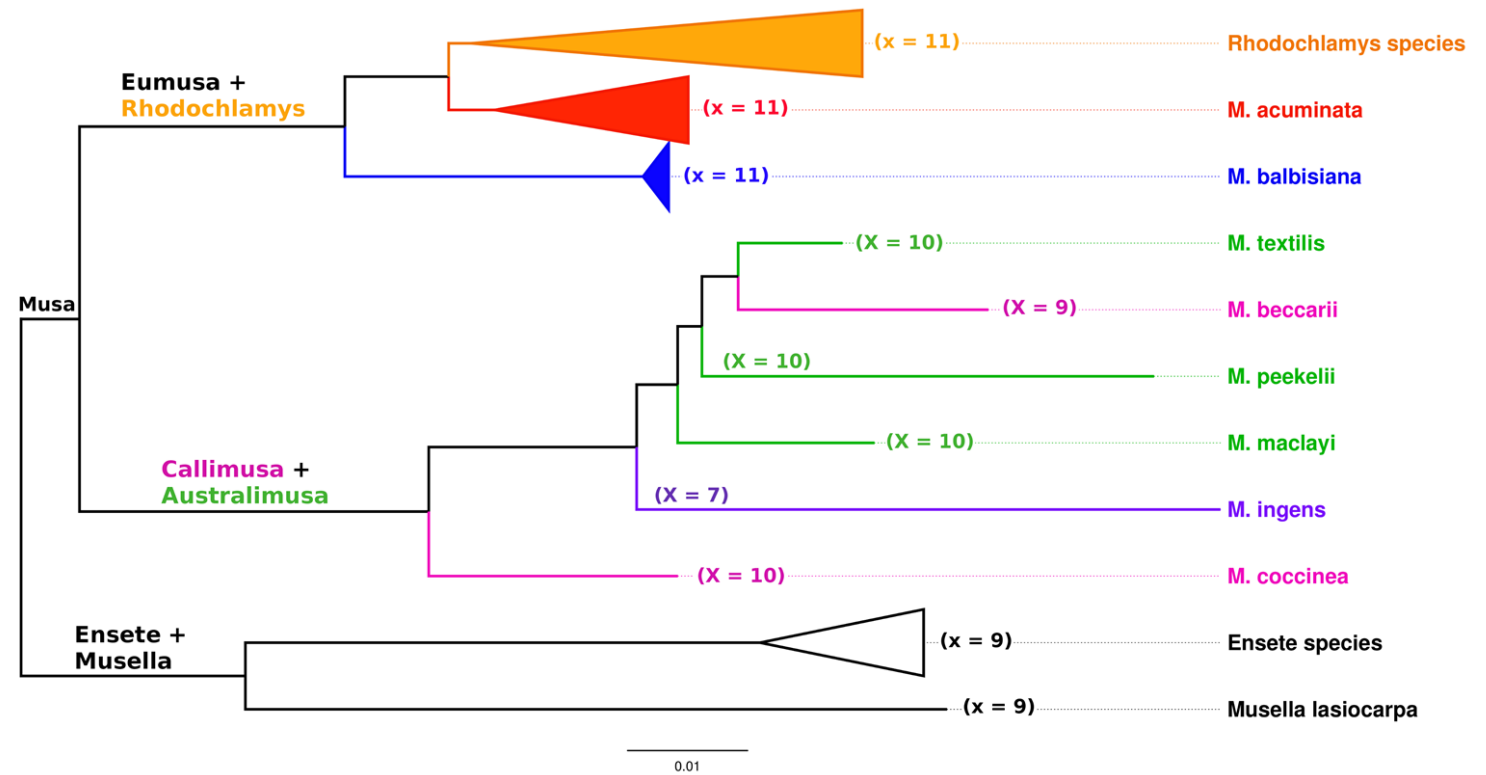
↓
Useful for introgression by chromosome-mediated gene transfer.





Examples from the genus *Musa*

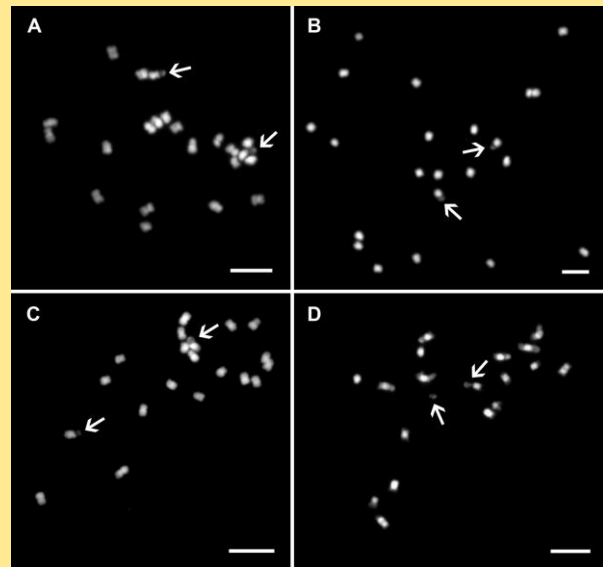
- Basic chromosome number, ploidy and genome size vary between closely related species
- Differences between karyotypes of species with the same ploidy and occurrence of structural chromosome heterozygosity



Cytogenomics methods

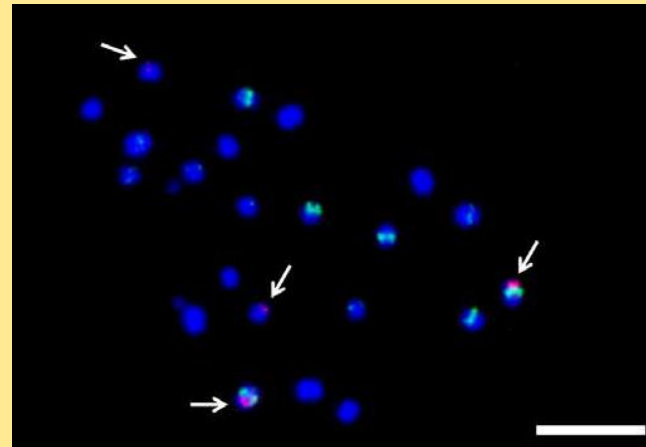
- Chromosome staining for determination of ploidy and chromosome number
- Molecular cytogenetics (fluorescence *in situ* hybridization – FISH) for detailed karyotype analysis

Chromosome counting

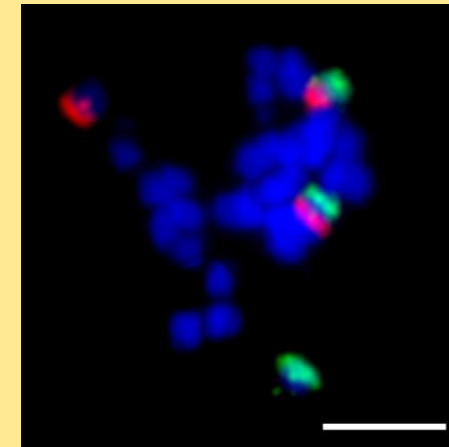


A) *Musa acuminata* ($2n=2x=22$); B) *Musa laterita* ($2n=2x=22$); C) *Musa maclayi* ($2n=2x=20$); D) *Musa beccarii* ($2n=2x=18$)
Arrows indicate NOR regions. Bar = 5 μ m

Molecular cytogenetics



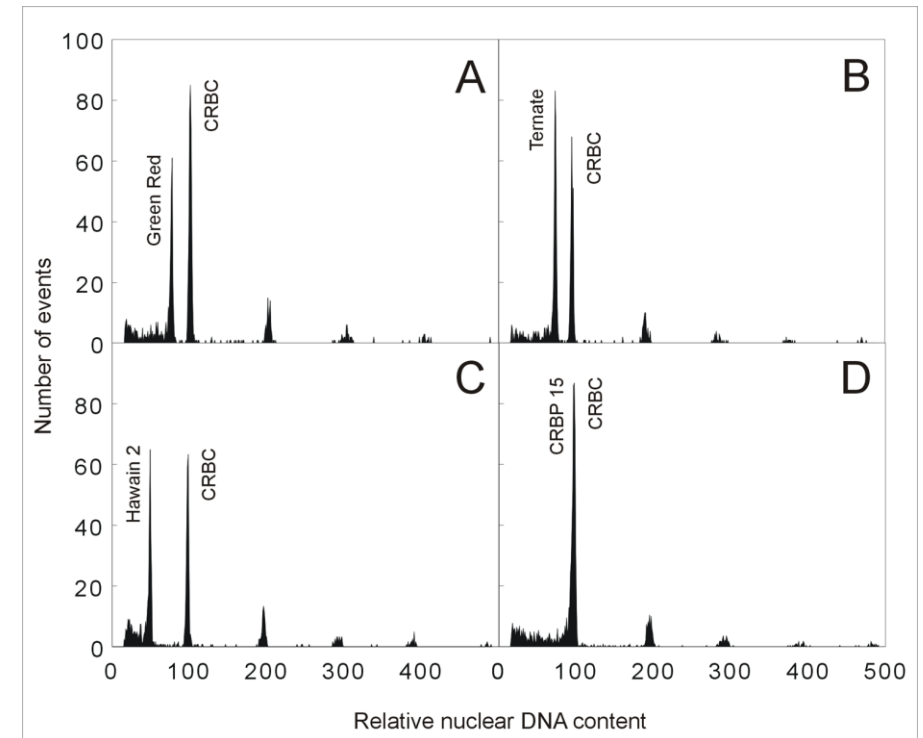
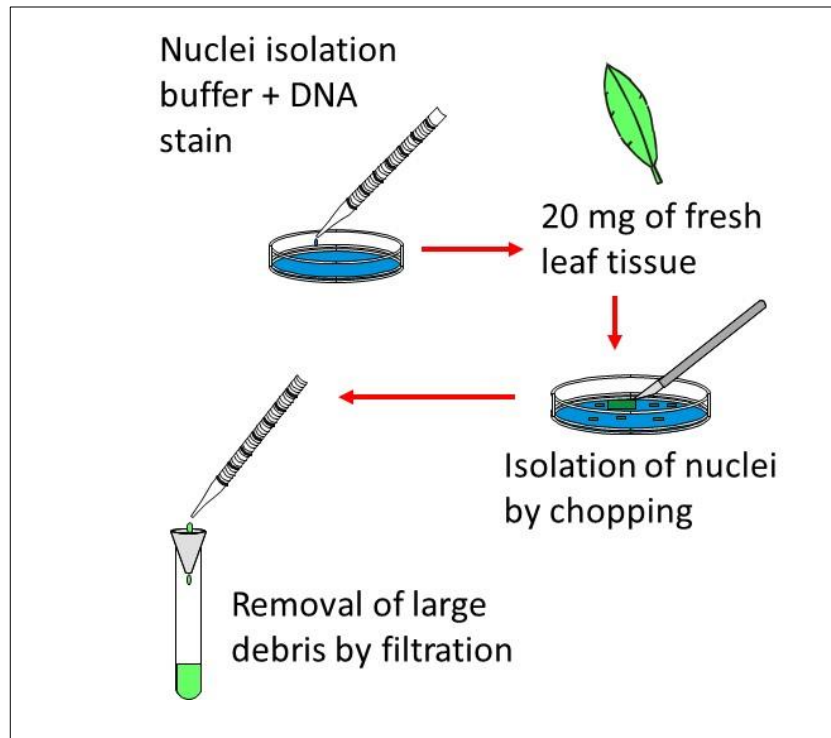
Fluorescence in situ hybridization (FISH) with probes to 5S rDNA (green) and satellite CL18 (red) in *Musa schizocarpa* ($2n=2x=22$). Chromosomes were counterstained with DAPI (blue). Bar = 5 μ m



Chromosome-arm specific oligo painting FISH in aneuploid clone 'Imbogo' ($2n = 3x - 1 = 32$). Long arm of chromosome 3 (green); short arm of chromosome 8 (red). Chromosomes were counterstained with DAPI (blue). Bar = 5 μ m

Ploidy/aneuploidy/genome size determination

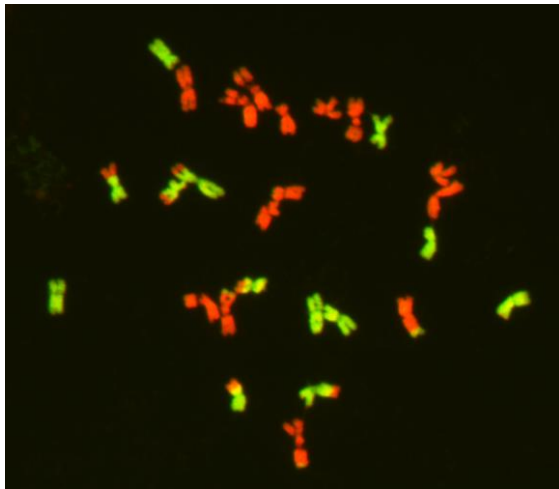
- Flow cytometry of nuclear DNA content as a rapid, accurate, easy and generally applicable method
- Characterization of >1500 accessions in *Musa* genebank (ITC collection at KU Leuven, Belgium)



Determination and facilitation of interspecific crossing/ introgression programs

- Genome composition of Festuca x Lolium hybrids and introgression lines
- Molecular cytogenetics (genomic *in situ* hybridization – GISH) for identification of parental chromosomes in interspecific hybrids and introgressions

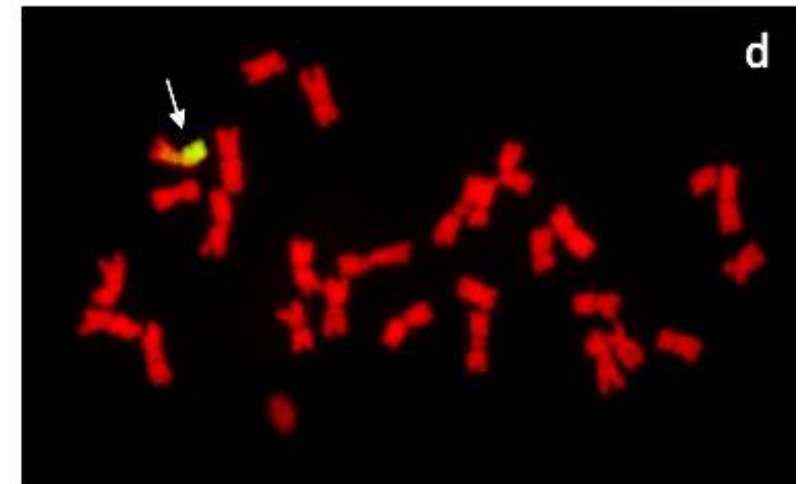
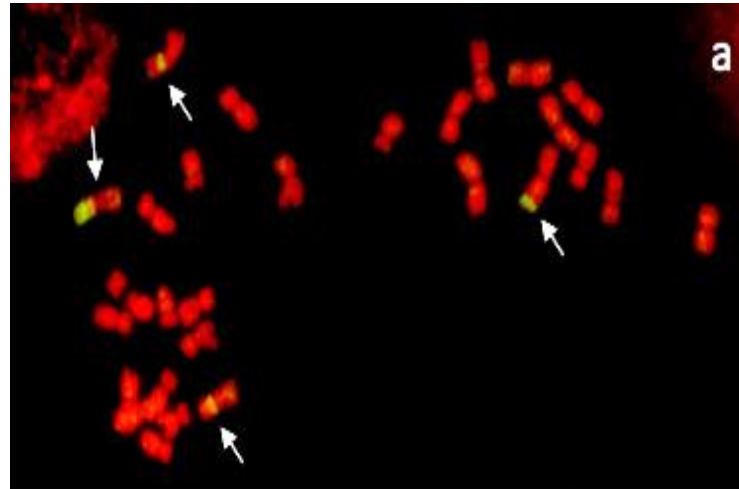
Festuca x Lolium hybrid



Fescue

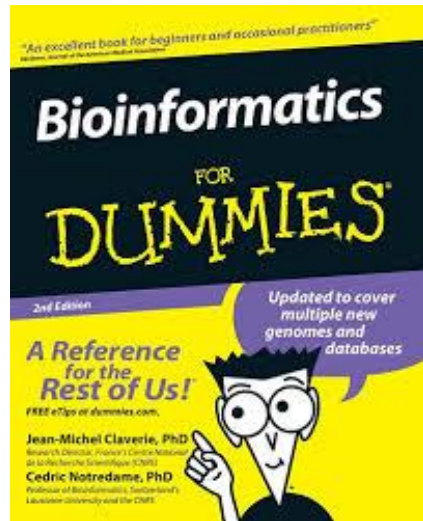
Ryegrass

Introgression lines



Bioinformatics

An interdisciplinary field of science that develops methods and software tools for understanding biological data.



It's not effortless!

PRO-GRACE aims to provide and refine bioinformatic methods for different purposes (D3.5)

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The screenshot displays the Galaxy web interface. The top navigation bar includes 'Galaxy', 'Workflow', 'Visualize', 'Shared Data', 'Help', 'User', and notification icons. A left sidebar contains tool categories: 'Tools' (with a search bar and 'Upload Data' button), 'Get Data', 'Send Data', 'Collection Operations', 'GENERAL TEXT TOOLS' (Text Manipulation, Filter and Sort, Join, Subtract and Group, Datamash), and 'GENOMIC FILE MANIPULATION' (FASTA/FASTQ, FASTQ Quality Control, SAM/BAM, BED, VCF/BCF). The main area is titled 'Published Workflows' and features a search bar, '+ Create' and 'Import' buttons, and a table of workflows.

Name	Tags	Updated	Owner
▼ QC report (imported from URL) ↗ rna-seq-reads-to-counts	transcriptomics	about 11 hours ago	abdul85
▼ Average Bigwig between replicates (release v0.1) We assume the identifiers of the input list are like: sample_name_replicateID. The identifiers of the output list will be: sample_name		5 days ago	iwc
▼ Repeat masking with RepeatModeler and RepeatMask (release v0.1)		5 days ago	iwc
▼ ATACseq (release v0.5.1) This workflow takes as input a collection of paired fastq. It will remove b ...	ATACseq	5 days ago	iwc

R Markdown

from R Studio

A collage of R Markdown content. It includes a map of Minnesota with 'Winnipeg' and 'James Bay' labeled, overlaid with mathematical formulas. A scatter plot shows 'mpg' vs 'hp' with a legend for 'am' (automatic/manual). A code block shows R code for ggplot2:

```
p <- ggplot(mtcars2, aes(hp, mpg, color = am)) +  
  geom_point()  
p  
p + geom_smooth()
```

Duplicates and misclassified

Based on SNPs identified using reduced representation/whole genome and chromosome scale (re)sequencing



Protocol will be developed/exploited to correct misidentifications, spot duplicates within and between collections



Exploit the barcode as well as a DOI associated to each of them, to minimize misidentifications and manage excessive levels of duplication

Duplicates and misclassified

the plant journal



Original Article | [Open Access](#) |

Analysis of >3400 worldwide eggplant accessions reveals two independent domestication events and multiple migration-diversification routes

Lorenzo Barchi , Giuseppe Aprea, M. Timothy Rabanus-Wallace, Laura Toppino, David Alonso, Ezio Portis, Sergio Lanteri, Luciana Gaccione, Emmanuel Omondi, Maarten van Zonneveld, Roland Schafleitner, Paola Ferrante, Andreas Börner, Nils Stein, Maria José Díez, Veronique Lefebvre, Jérémy Salinier, Hatice Filiz Boyaci, Richard Finkers, Matthijs Brouwer, Arnaud G. Bovy, Giuseppe Leonardo Rotino, Jaime Prohens, Giovanni Giuliano ... [See fewer authors](#) ^

First published: 08 September 2023 | <https://doi.org/10.1111/tpj.16455>

Our approach: calculate identity by state (IBS) in pair-wise comparisons

Which threshold to define accessions as duplicates?

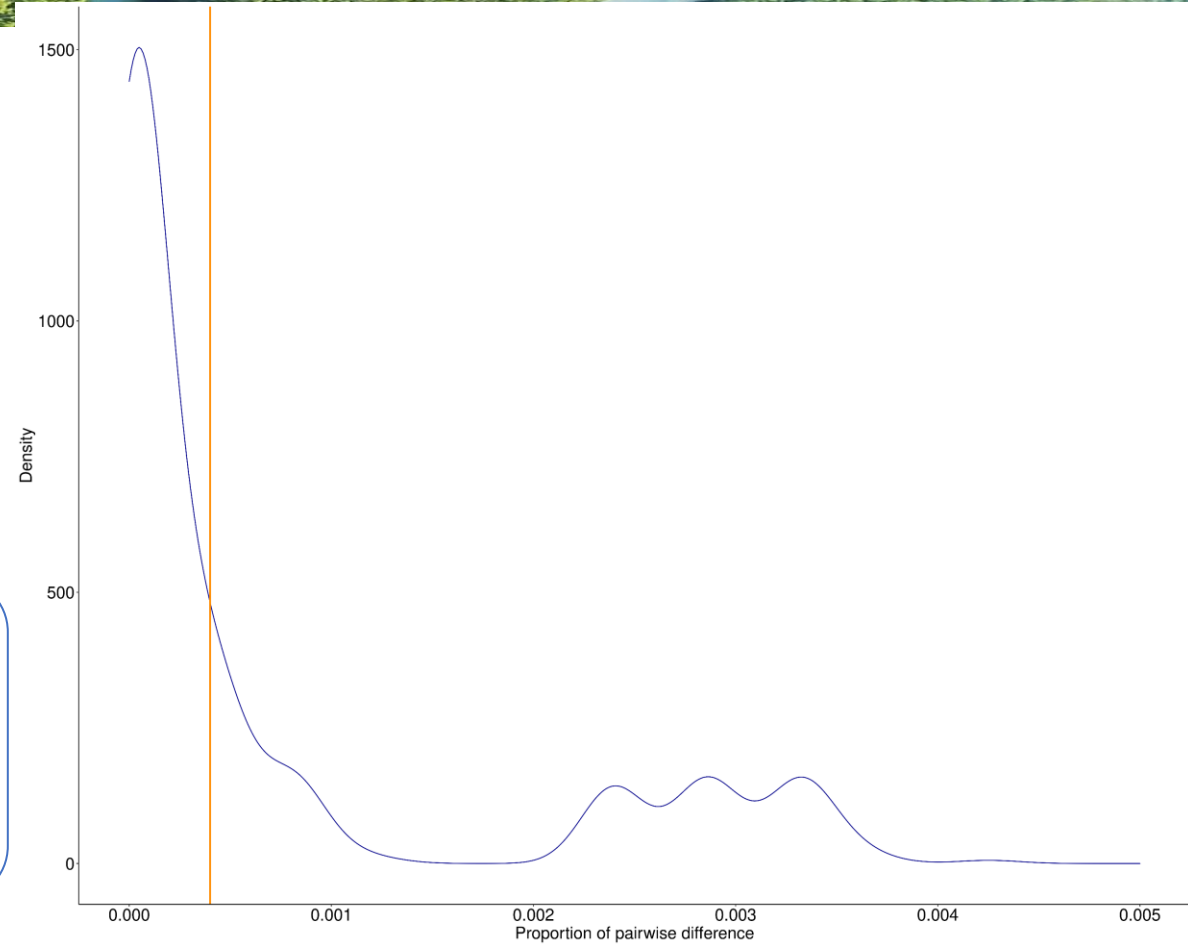


Used replicates, i.e. additional samples from a well-known accession

Duplicates and misclassified



ML tree clearly shows identity among replicates



The distribution on controls IBs allowed to impose a cut-off value of dissimilarity ≤ 0.0005

Duplicates and misclassified

- For misclassification detection, for each accession, we determined the set of the 10 nearest neighbours and verified the species they belong to.
- A potential misclassification was called when the species match rate was less than 35%.
- The putative mislabeled accessions were eventually re-assigned to another species according to:
 - (i) assigned to a group having at least six representatives of another speciesand
 - (ii) after manual curation based on genebank passport data.

CWRs are source of genes for resistance to biotic and abiotic stresses exploitable for domesticated species breeding



Critical gaps in *ex situ* and *in situ* collections with respect to natural genetic variability of a species have been identified.

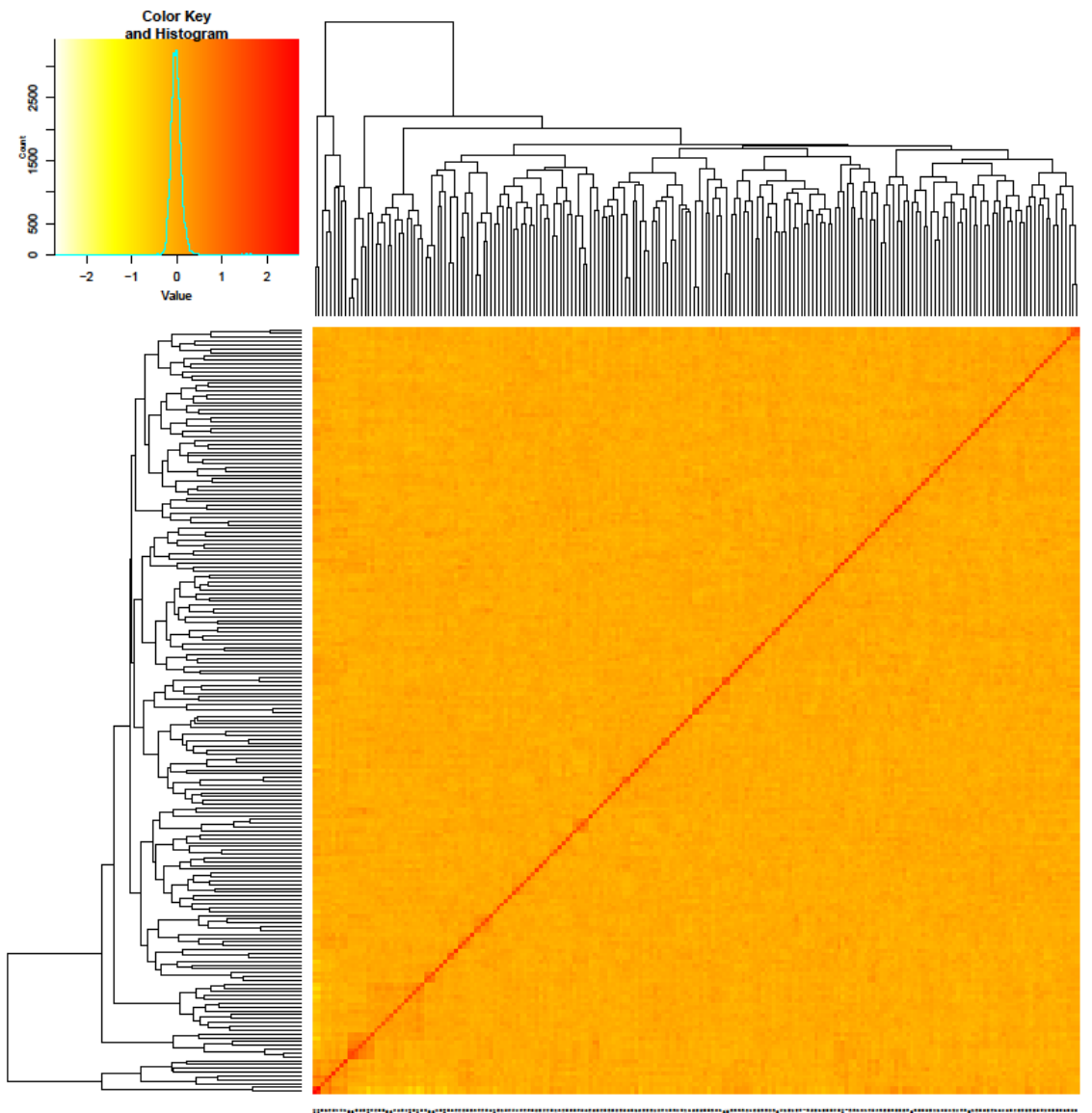
CWR are also poorly represented in *ex situ* and *in situ* collections



Procedure for population structure and kinship estimates will be provided. Together with barcoding, this will help to assist gap analysis and to reconstruct missing relations

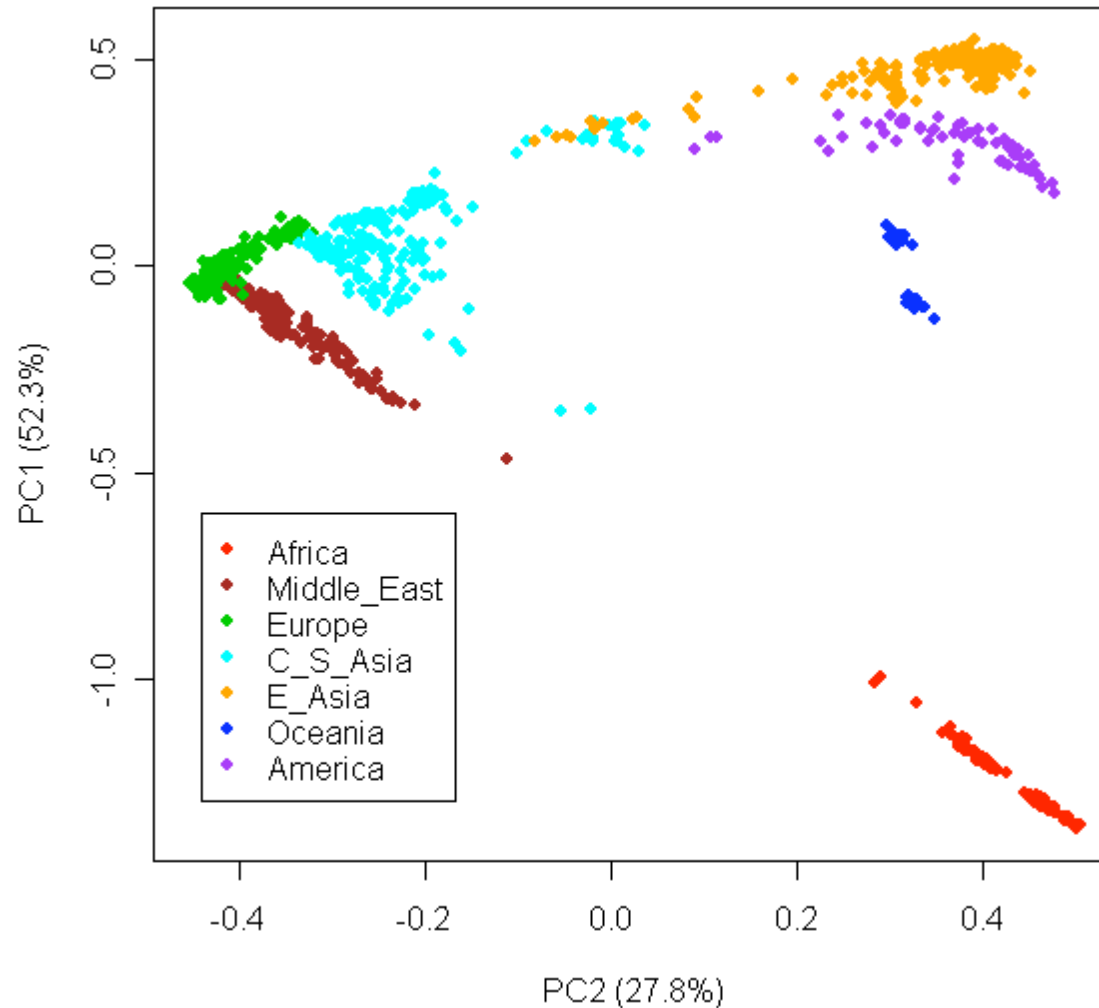
Kinship

Genetic resemblance between individuals



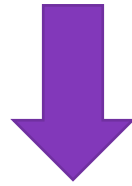
Population stratification

Population stratification is the presence of a systematic difference in allele frequencies between subpopulations in a population due to different ancestry



Core collection

Exploiting diversity/population structure/pedigree information



Construct core collections representative of the genetic variation of a much larger genepool



Set up pipelines for QTL and GWA studies which are now routinely used in the agronomic evaluation of large collections of germplasm

Build up the eggplant core collection

SNPs



VCF



Distance Matrix

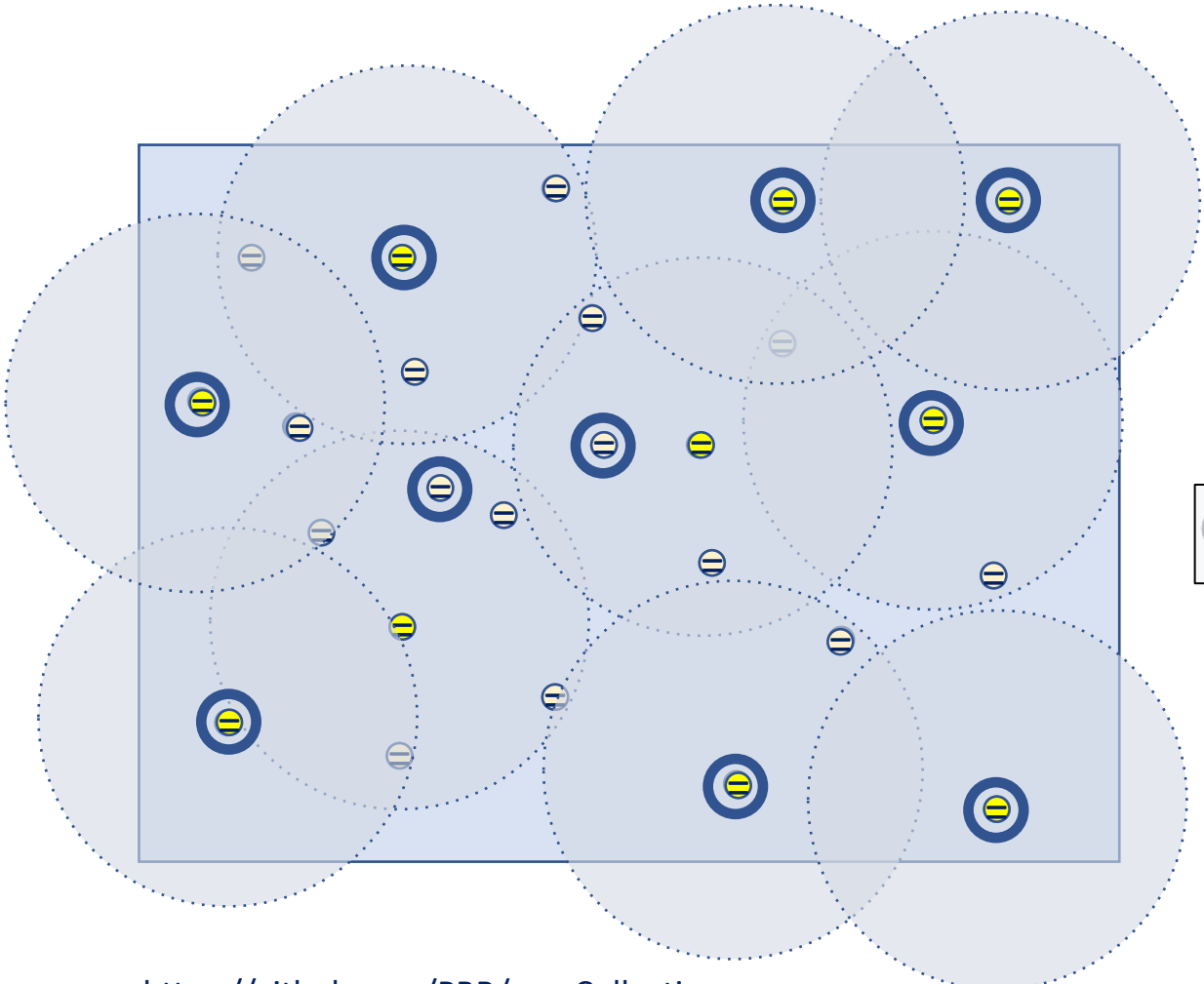
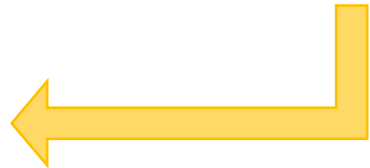


CoreCollection algorithm

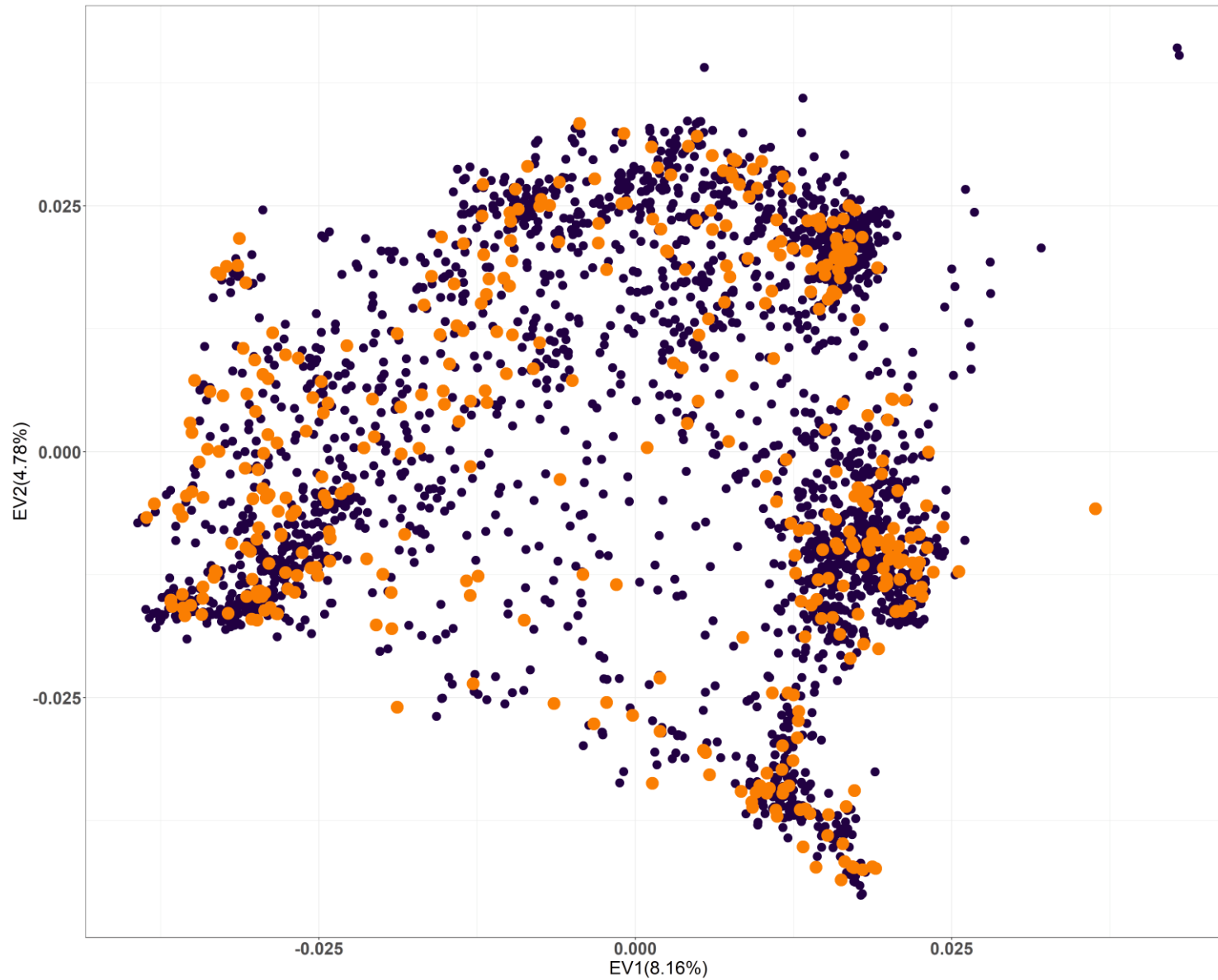


Core Collection

Preselected individuals

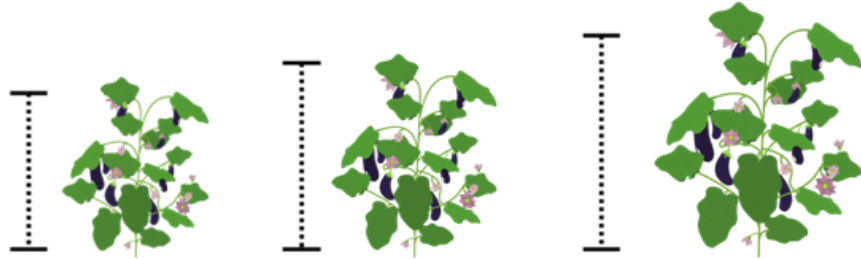


Build up the eggplant core collection



QTL and GWA

Phenotype



Genotype



SNPs, k-mers and SVs



Genomic Association and Prediction Integrated Tool

(Version 3)

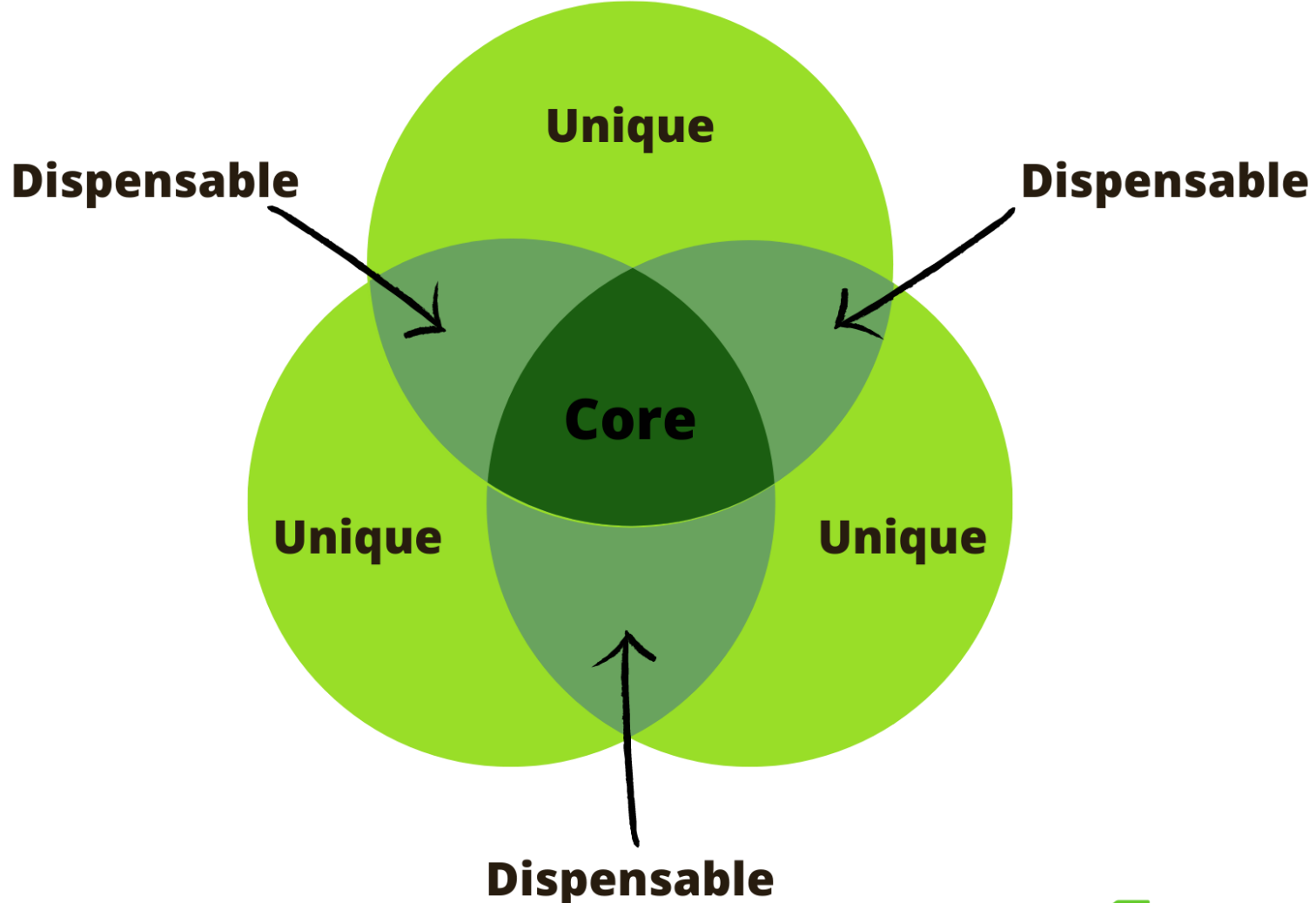
Pangenome construction can provide a comprehensive picture of the allelic variation within a single or among multiple species



Provide methodologies for pangenome construction, graph-pangenome, SNPs and indels identification as well as presence-absence variants (PAVs) and Selective Sweeps identification linked to key traits



Pan-genome



Eggplant reseq Illumina 20X

- Reference eggplant
inbred line '67/3'

- 23 *Solanum melongena*

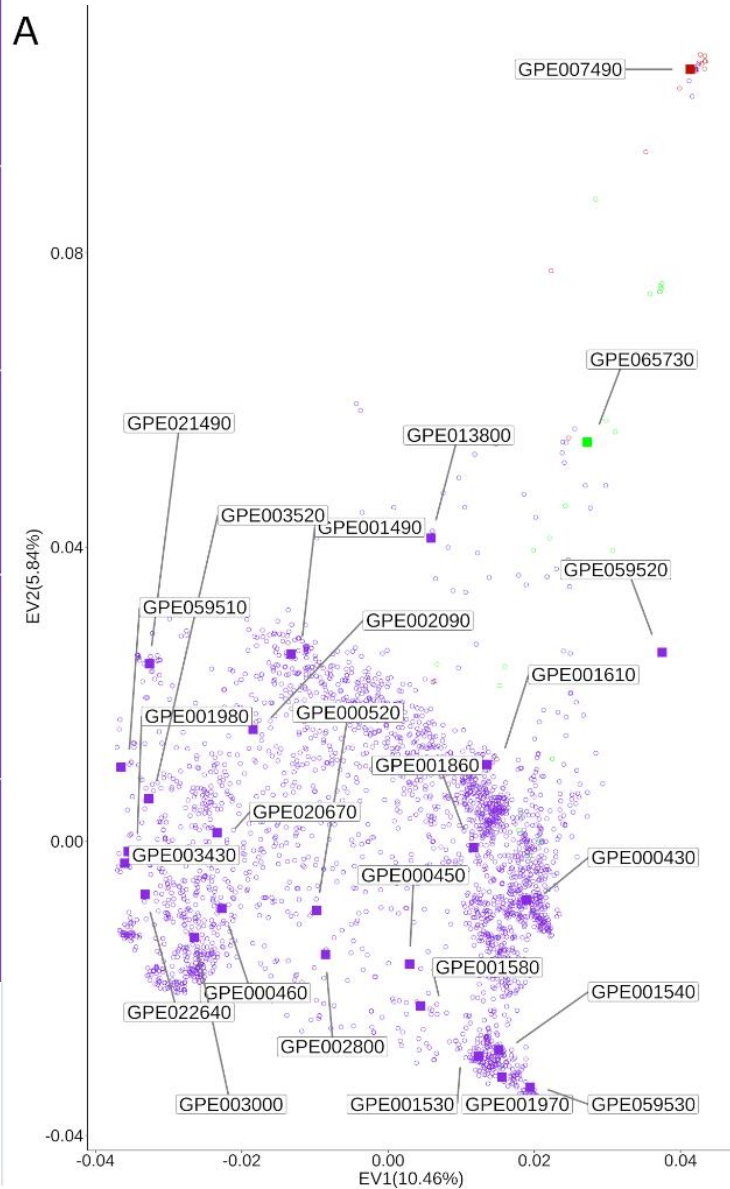
representative of the worldwide
phenotypic and genetic diversity
of the species

- 1 *Solanum insanum*

- 1 *Solanum incanum*



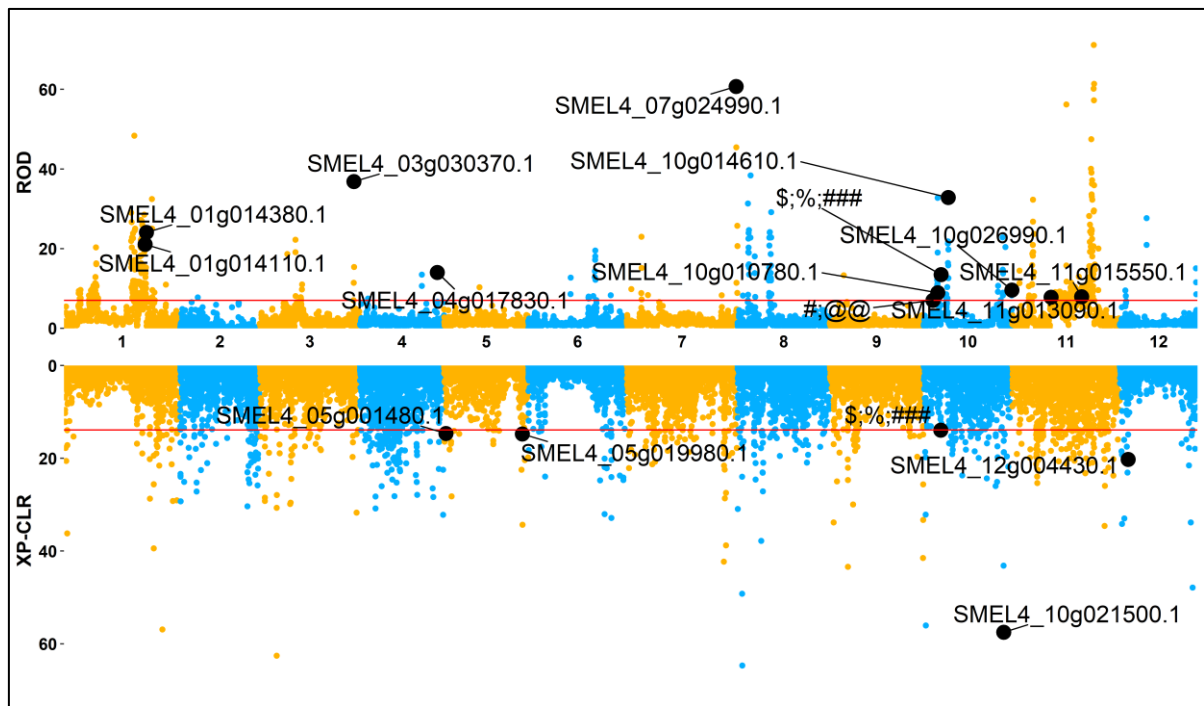
A



Selective sweeps and domestication

53 selective sweeps (spanning 35.3 Mb) identified

Non-anthocyanic vs anthocyanic fruits



Candidate genes identified in the SS regions.

Homologs of :

- Anthocyanin biosynthetic genes
FLAVONOL SYNTHASE (FLS)
CHALCONE ISOMERASE (CHI)
ANTHOCYANIDIN REDUCTASE (ANR)
DIHYDROFLAVONOL 4-REDUCTASE (FRR)

Positions of previously discovered QTLs and QTNs

- MATE and ABC transporters mediating anthocyanin sequestration in other species
 \$ Baren et al., 2012;
 ### Cericola et al., 2014;
- Prx31 peroxidase involved in anthocyanin degradation
 % Portis et al., 2014;
 # Frary et al., 2014;
- Transcription factors involved anthocyanin / proanthocyanidin regulation such as MYB14 and SmMYB1
 @ @ Toppino et al., 2016;
 @ @ Wei et al., 2020a;
 PI locus from Miyatake et al., 2020

Several **QTLs** and **QTNs** controlling fruit anthocyanin pigmentation **co-localized with the SS on chr. 10**



PRO-GRACE

PROMOTING A PLANT GENETIC
RESOURCE COMMUNITY FOR EUROPE



**Thank you for
your attention**